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# Stimulation of Phenotypically Normal Shoot Regeneration of Tomato (*Lycopersicon* esculentum Mill.) by Commercial Filter Paper Extract

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#### Summary

Effects of extracts from filter paper on the shoot regeneration of tomato (*Lycopersicon* esculentum Mill.) were examined. Cotyledon segments of tomato were cultured on supporting material made of polyester supplemented with Murashige and Skoog liquid medium, containing  $0.1 \text{ mg} \cdot \text{liter}^{-1}$  indole-acetic acid,  $1.0 \text{ mg} \cdot \text{liter}^{-1}$  zeatin and 3% sucrose. Soluble components were extracted from filter paper with hot water and added to the medium. Phenotypically normal shoot regeneration was stimulated by the extracts of filter paper; the highest percentage of shoot regeneration was obtained by an extract equivalent to 1 g filter paper per ml. When filter paper extracts were fractionated on Sephadex G-25 gel column, the shoot regeneration stimulating activity was found in a low molecular weight fraction. These results suggest that filter paper contains thermostable substances low molecular weight which induce shoot regeneration.

Key Words: biological activity, filter paper, polyester support, shoot regeneration, tomato.

#### Introduction

Filter paper is a material used for removing solids from solutions, support for paper chromatography and a moist medium for germinating seeds. Furthermore, filter paper is often applied for paper-bridge-(Bhat et al., 1992) and paper-wick-systems (Baker and Phillips, 1962) in plant tissue culture.

Ichimura et al. (1995) reported that gelling agents or supporting materials, derived from plant polysaccharides, were much more effective for shoot regeneration from cotyledonary segments of tomato than any other support materials made from polyester or ceramic. Furthermore, Ichimura and Oda (1995) and Ichimura et al. (1995) found that wood pulp and agar extracts stimulated shoot regeneration of tomato when they were added to a liquid medium utilizing a polyester support. These results suggest that some water-soluble substances derived from plant polysaccharides promote shoot regeneration of tomato. Filter paper is made primarily of cellulose from cotton; its extract may have some stimulating agents for shoot regeneration.

The purpose of this study is to investigate effects of filter paper extracts on adventitious shoot regeneration from tomato tissues.

## **Materials and Methods**

Nine hundred g of filter paper (No. 2, Toyo Roshi Kaisha, Ltd.) was minced (ca. 2 cm diam.) and soaked

in 18 liter of distilled water at a temperature of 80 °C. The mixture was kept at 80 °C in an oven for 1 hr. Upon cooling to room temperature, the mixture was filtered through the same type of filter paper and concentrated to 140 ml *in vacuo* with a rotary evaporator below 50 °C. The concentrated extract was frozen at -30 °C until it was used.

Supporting material made of polyester (Cloud; Toyobo Co., Ltd.) was placed into culture vessels (6.2  $\times$  6.2  $\times$  9.8 cm). The weight and size of polyester support were 1.5 g and 5.5 $\times$ 5.0 $\times$ 1.5 cm, respectively. MS liquid medium (Murashige and Skoog, 1962) supplemented with 3% sucrose, 0.1 mg  $\cdot$  liter<sup>-1</sup> indole-acetic acid and 1.0 mg  $\cdot$  liter<sup>-1</sup> zeatin and a known amount of filter paper extracts (pH 5.8), was poured into the vessel and autoclaved at 121 °C for 10 min. The total volume of medium was 55 ml. Four excised cotyledonary segments (ca. 5 $\times$ 3 mm) of tomato (*Lycopersicon esculentum* Mill. cv. Zuiken) prepared according to Ichimura and Oda (1995) were placed onto the support in each of two vessels per treatment. The segments were cultured as described by Ichimura et al. (1995).

To examine molecular weights of active substances, an extract obtained from filter paper (7 kg) was concentrated to 43 ml as above. The concentrate was loaded onto a Sephadex G-25 (fine) column ( $4 \times 90$  cm); which was equilibrated and then eluted with distilled water at room temperature. The flow rate was 100 ml  $\cdot$  hr<sup>-1</sup>; 12 ml in each fraction was collected. The column was calibrated with blue dextran (mol. wt. 2,000,000), stachyose tetrahydrate (mol. wt. 738), sucrose (mol. wt. 342), and glucose (mol. wt. 180). Aliquots of each fraction were assayed for sugar by the method of Dubois et

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# al. (1956).

#### **Results and Discussion**

In this study, we used polyester support instead of agar, because polyester support is guaranteed not to release its contents into the medium nor to absorb components from it (Toyobo, 1988), so that, only the direct effects of medium on explants can be observed. Irrespective of presence of filter paper extracts, margins of the explants became callused after 2 weeks of culture, and adventitious buds regenerated from the calluse after 3 weeks. There was little difference in the morphology of adventitious buds between tissues exposed to filter paper extracts and those of the control up to seven weeks of culture. Thereafter, differences in the subsequent growth between the treatment and control became apparent. Most adventitious shoots in the control tissues became abnormally thickened and frequently vitrified and phenotypically normal shoots rarely regenerated. The addition of the filter paper extracts to the polyester medium stimulated the adventitious buds to grow into phenotypically normal shoots. This result reveals that the filter paper extracts contain substances that stimulate phenotypically normal shoot regeneration. The optimum concentration of the extracts for shoot growth was calculated to be equivalent to 1 g per ml (Table 1). The weight of the extracts recovered from 1,000 g of filter paper being about 280 mg, the concentration of the extract equivalent to 0.1 g of filter paper per liter which was the minimum concentration to stimulate shoot regeneration is estimated to be 0.3 mg  $\cdot$  liter<sup>-1</sup>. Hence, some stimulating substances contained in the commercial filter paper is effective at a concentration less than one ppm level.

The filter paper extract was fractionated on a Sephadex G-25 column, and the sugar concentration in each fraction of the eluate was assayed because filter paper mainly consisted of the plant polysaccharide, cellulose. Whereas the main component of the extract was

a polysaccharide-like substance, a low molecular weight fraction was also positive to phenol-sulfuric acid reaction (Fig. 1). We divided the eluate into 4 fractions, and examined the effects of fractions on shoot regeneration. Percentages of explants with normal leaves, number and length of normal leaves exposed to fraction III whose molecular weight was equivalent to several hundreds were greater than those exposed to any other fractions (Table 2). These results indicate that some stimulating substances are a low molecular weight compound.

To our knowledge, we are first to report that commercial filter paper has biological activity. This finding should be of interest because filter paper is routinely used in biological and chemical experiments, especially in tissue culture as a paper-bridge-system which facili-



Fig. 1. Sephadex G-25 elution profile of filter paper extract. Twelve ml in each fraction was collected and aliquot (10  $\mu$ l) of each column fraction was assayed for sugar content. Curve represents sugar content expressed as absorbance at 490 nm. Void volume (Vo) was determined by blue dextran. Elute portions of stachyose tetrahydrate (ST), sucrose (SU) and glucose (GL) were indicated by arrows. Eluate was divided into 4 fractions, I, II III and IV, and their effects on the shoot regeneration was shown in Table 2.

**Table 1.** Effect of filter paper extract on shoot regeneration of tomato. The data were taken after 104days of culture. Values represent mean of 8 replications  $\pm$  standard errors.

| Concentration                      | Percentage<br>of explant                  | Number of<br>shoot per<br>explant <sup>y</sup> | Number of<br>shoot with                   | Shoot<br>length <sup>w</sup><br>(mm) | shoot length<br>with normal<br>leaves <sup>°</sup><br>(mm) |
|------------------------------------|-------------------------------------------|------------------------------------------------|-------------------------------------------|--------------------------------------|------------------------------------------------------------|
| (g equivalent · ml <sup>-1</sup> ) | with normal<br>leaves <sup>2</sup><br>(%) |                                                | normal leaves<br>per explant <sup>*</sup> |                                      |                                                            |
| 0                                  | 12.5                                      | $0.6 \pm 0.2$                                  | 0.1 ± 0.1                                 | $11.0 \pm 3.6$                       | 4.5 ± 4.2                                                  |
| 0.001                              | 50                                        | $1.1 \pm 0.5$                                  | $0.6 \pm 0.3$                             | $18.6 \pm 7.9$                       | $16.0 \pm 8.5$                                             |
| 0.01                               | 50                                        | $1.6 \pm 0.4$                                  | $0.9\pm0.4$                               | $29.4 \pm 10.0$                      | $24.6 \pm 11.2$                                            |
| 0.1                                | 75                                        | $1.0 \pm 0.2$                                  | $0.8 \pm 0.2$                             | $23.6~\pm~8.6$                       | $22.0~\pm~~9.0$                                            |
| 1                                  | 100                                       | $1.4 \pm 0.3$                                  | $1.3 \pm 0.3$                             | $44.9 \pm 12.2$                      | $44.5\pm12.4$                                              |
| 2                                  | 87.5                                      | $0.8\pm0.3$                                    | $0.8\pm0.3$                               | $27.4 \pm 17.2$                      | $27.4 \pm 17.2$                                            |

<sup>4</sup> Percentage of explant produced phenotypically normal leaves.

<sup>y</sup> Number of shoots longer than 10 mm.

<sup>x</sup> Number of shoots longer than 10 mm with phenotypically normal leaves.

\* Length of longest shoot each explant produced.

<sup>v</sup> Length of longest shoot with phenotypically normal leaves each explant produced.

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Table 2. Effect of fractions of filter paper extract separated by aSephadex G-25 column on phenotypically normal shootregeneration of tomato. Values represent mean of 8 replica-tions. The data were taken after 105 days of culture.

| Fraction | Concentration<br>(g equivalent<br>·ml <sup>-1</sup> ) | Percentage<br>of explant<br>with normal<br>leaves <sup>2</sup><br>(%) | Number of<br>shoot with<br>normal leaves<br>per explant <sup>y</sup> | Shoot length<br>with normal<br>leaves <sup>x</sup><br>(mm) |
|----------|-------------------------------------------------------|-----------------------------------------------------------------------|----------------------------------------------------------------------|------------------------------------------------------------|
| Control  | 0                                                     | 0                                                                     | 0                                                                    | 0                                                          |
| I        | 0.22                                                  | 0                                                                     | 0                                                                    | 0                                                          |
|          | 2.2                                                   | 0                                                                     | 0                                                                    | 0                                                          |
| II       | 0.22                                                  | 0                                                                     | 0                                                                    | 0                                                          |
|          | 2.2                                                   | 0                                                                     | 0                                                                    | 0                                                          |
| III      | 0.22                                                  | 50.0                                                                  | 0.6                                                                  | 24.8                                                       |
|          | 2.2                                                   | 37.5                                                                  | 0.6                                                                  | 23.5                                                       |
| IV       | 0.22                                                  | 25.0                                                                  | 0.1                                                                  | 12.8                                                       |
|          | 2.2                                                   | 12.5                                                                  | 0.1                                                                  | 1.5                                                        |

<sup>2</sup> Percentage of explant produced phenotypically normal leaves.

<sup>y</sup> Number of shoots longer than 10 mm with phenotypically normal leaves.

<sup>x</sup> Length of longest shoot with phenotypically normal leaves each explant produced.

tated rooting (Bhat et al., 1992), and the paper-wicksystem for apical meristem culture (Baker and Phillips, 1962) and floral differentiation of *Torenia* (Kobayashi et al., 1993). These positive effects on cultured tissues may also be related to the shoot regeneration-stimulating substances in filter paper.

The regeneration-stimulating activity was partitioned by hot water extracts, so the stimulant seems to be a heat-stable, hydrophilic compound. Agar or supporting materials made of wood pulp or cotton fiber or their extracts were found to stimulate shoot regeneration of tomato (Ichimura and Oda, 1995; Ichimura et al., 1995). Therefore, the active substance in filter paper may be related to plant polysaccharides just as Bois (1992) suspected because, in that study, cotton hydrolysate induced differentiation and organogenesis from callus of strawberry.

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市販ろ紙の抽出液によるトマト子葉からのシュート再生の促進

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### 摘要

市販ろ紙の熱水抽出液がトマト子葉片からのシュート再生 に及ぼす影響について調べた.トマト子葉片をポリエステル 製の支持体上で培養したところ,ろ紙抽出液によりトマト子 葉片からのシュート再生は著しく促進された.抽出液を Sephadex G-25 カラムで分画したところ,主要な成分は高

分子画分にみられたが,活性は低分子画分に存在した.以上 の結果より,市販のろ紙にはトマトのシュート再生を促進す る熱安定性の低分子の物質が含まれていることが明らかとなった.

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