

Triterpenoids from Flower Buds of *Tussilago farfara* L.

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A new triterpenoid, bauer-7-ene-3 β ,16 α -diol, and two known triterpenoids, bauerenol and isobauerenol, were isolated from dried flower buds of *Tussilago farfara* L. (Compositae). The structure of the new compound was elucidated by the spectroscopic data.

Keywords : *Tussilago farfara* ; flower bud; Compositae; triterpenoid

The flower buds of *Tussilago farfara* L. (Compositae) have been widely used for treatment of cough, bronchitis and asthmatic disorders in China.¹⁾ In previous papers, we reported the isolation and structural elucidation of the essential oil components²⁾ and sesquiterpenoids³⁾ from the plant. In the present paper, we report the isolation and structural elucidation of a new triterpenoid, bauer-7-ene-3 β ,16 α -diol (**1**). Two known compounds, bauerenol (**2**)⁴⁾ and isobauerenol (**3**)⁵⁾ were also isolated, and identified by its spectral data. This is the first report of isolation of **2** and **3** from the flower buds of *T. farfara*. Extraction and isolation were carried out as described in the experimental section.

Compound **1** was shown to have the molecular formula $C_{30}H_{50}O_2$ by high-resolution (HR)-MS data. The IR spectrum showed the presence of hydroxyl groups (3329 cm^{-1}), and the electron ionization (EI)-MS gave significant fragment ion peaks at m/z 424 ($M^+ - H_2O$), 406 ($M^+ - 2H_2O$), 273 (a), 255 (a - H_2O), 247 (b), 229 (b - H_2O), 221 (c) and 203 (c - H_2O) (Fig. 1). These fragment ions suggested that **1** was a bauerane or multiflorane derivative with hydroxyl groups in the rings A/B and rings D/E of the molecule.⁶⁾ The 1H -NMR spectrum of **1** indicated the presence of six tertiary methyl groups, two secondary methyl groups, two hydroxy methine groups and one trisubstituted olefinic proton. The 1H -detected heteronuclear multiple bond connectivity (HMBC) analysis showed correlations between H_3 -24 and C-3, C-4 and C-5; H_3 -25 and C-1, C-

5, C-9 and C-10; H_3 -26 and C-8, C-13 and C-14; H_3 -28 and C-16, C-17, C-18 and C-22; H_3 -29 and C-18, C-19 and C-20; and H_3 -30 and C-19, C-20 and C-21, suggesting that **1** was a bauerane-type triterpenoid (Fig. 2). The HMBC spectrum determined the position of the trisubstituted double bond hydrogen to be at C-7, by showing correlations between H-7 and C-5 and C-9; and H_3 -26 and C-8, and the positions of the hydroxyl groups to be at C-3 and C-16, by the correlations between H_3 -24 and C-3; H-15 and C-16; and H_3 -28 and C-16 (Fig. 2). From the above data, the planar structure of **1** was determined to be bauer-7-ene-3,16-diol. The stereostructure was ascertained by the nuclear Overhauser effect (NOE) interactions observed in the nuclear Overhauser enhancement spectroscopy (NOESY) spectrum (Fig. 3). These NOEs implied *trans*-junctions of the rings A/B and the rings C/D, and *cis*-junction of the rings D/E, and that the rings A and D had chair conformation, the ring B half chair conformation and the rings C and E boat conformation. The configuration of the hydroxyl groups at C-3 and C-16 were determined by 1H -NMR and NOESY spectra. In the 1H -NMR spectrum, the coupling patterns and constants for H-3 [δ_H 3.21 (dd, $J=11.0, 4.8Hz$)] and H-16 [δ_H 3.62 (dd, $J=12.8, 4.8Hz$)] suggested that the hydroxyl groups at C-3 and C-16 had β and α configurations, respectively. The same conclusion was derived from the NOESY cross-peaks observed between H-3 α and H_3 -23; and H-16 β and H_3 -26 (Fig. 3). Accordingly, the structure of **1**

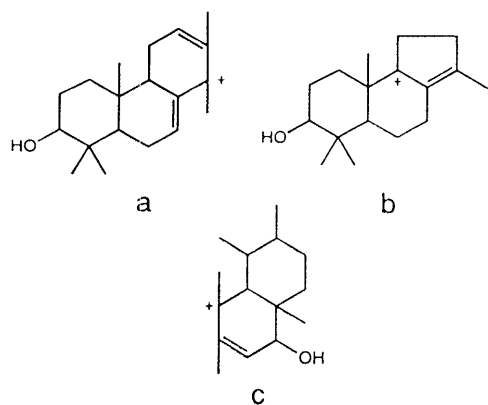


Fig. 1

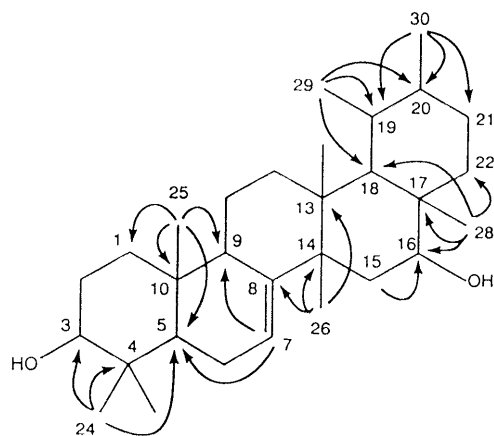


Fig. 2

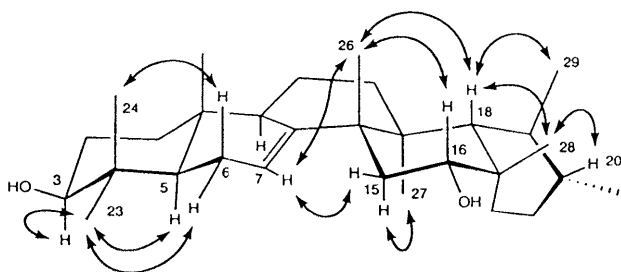


Fig. 3

was determined to be bauer-7-ene-3 β ,16 α -diol.

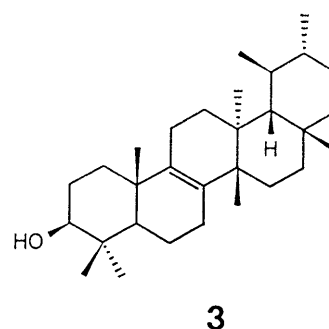
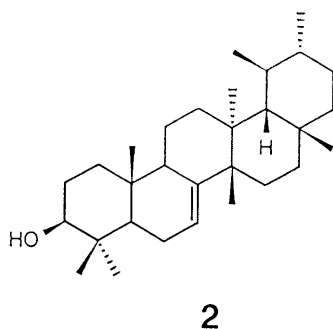
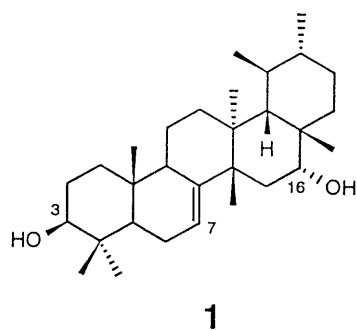
EXPERIMENTAL

General Procedure Melting point was determined on a Yanagimoto micromelting point apparatus and are uncorrected. Optical rotations were determined with a JASCO DIP-360 digital polarimeter. IR spectra with a Parkin-Elmer FT-IR 1725X infrared spectrophotometer, ^1H - and ^{13}C -NMR spectra with a JEOL JNM LA 600, JNM LA 400, JNM EX 270 spectrometer and EI- and HR-MS on a JEOL JMS-DX 303 mass spectrometer. Column chromatography was carried out on Kieselgel 60 (Merck: 230 - 400 mesh) and preparative HPLC on a Tosoh HPLC system (pump; CCPD, detector; RI-8010).

Plant material Dried buds of *Tussilago farfara* were purchased from Tochimoto Tenkaido Co., Ltd., Osaka, Japan in 1990.

Extraction and isolation The dried buds of *T. farfara* (5.0 kg) were extracted with Et_2O at room temperature for one week. The Et_2O extract was subjected to steam distillation to give an essential oil (7.7 g) and residue (64.5 g). A part of this residue (23.5 g) was placed on a silica gel column and developed with benzene-AcOEt (9:1). The eluate was separated into 17 fractions (frs.1 - 17). Fr. 9 was purified by preparative HPLC (column; TSK gel ODS-120T, 7.8mmi.d.x 30cm; mobile phase, MeOH; column temperature, 40 $^\circ\text{C}$; flow rate, 1.0 ml/min; RI detector) to give **2** (1.4 mg) and **3** (1.9 mg). Fr. 15 was recrystallized from CHCl_3 to give **1** (1.6 mg).

Bauer-7-ene-3 β ,16 α -diol (1) Colorless needles, mp. 282-285 $^\circ\text{C}$. $[\alpha]_D^{28}$ -25.6 $^\circ$ ($c=0.2$, pyridine). IR $_{\text{max}}$ (KBr) cm^{-1} : 3329. HR-MS m/z : 442.3788 (M^+ , Calcd for $\text{C}_{30}\text{H}_{50}\text{O}_2$; 442.3811). EI-MS m/z : 442 (M^+), 424 ($\text{M}^+ - \text{H}_2\text{O}$), 406 ($\text{M}^+ - 2\text{H}_2\text{O}$), 273 (a), 255 (a - H_2O), 247 (b), 229 (b - H_2O), 221 (c), 203 (c - H_2O). ^1H -NMR ($\text{CDCl}_3 + \text{CD}_3\text{OD}$ (4:1), 600MHz) δ : 0.76 (3H, s, H_3 -25), 0.85 (3H, s, H_3 -24), 0.93 (3H, d, $J=6.2\text{Hz}$, H_3 -30), 0.96 (3H, s, H_3 -27), 0.97 (3H, s, H_3 -23), 1.04 (3H, s, H_3 -26), 1.08 (3H, d, $J=7.0\text{Hz}$, H_3 -29), 1.22 (3H, s, H_3 -28), 1.31 (2H, m, H-5, H-18), 1.53 (1H, dd, $J=12.8, 12.8\text{Hz}$, H-15 α), 1.57 (1H, m, H-20), 1.71 (dd, $J=12.8, 4.8\text{Hz}$, H-15 β), 1.98 (1H, m, H-6 β), 2.14 (1H, m, H-6 α), 3.21 (1H, dd, $J=11.0, 4.8\text{Hz}$, H-3), 3.62 (1H, dd, $J=12.8, 4.8\text{Hz}$, H-16), 5.45 (H, d, $J=3.7\text{Hz}$, H-7). ^{13}C -NMR ($\text{CDCl}_3 + \text{CD}_3\text{OD}$ (4:1), 100MHz) δ : 12.96 (C-25), 14.68 (C-24), 16.73 (C-11),



22.66 (C-30), 22.67 (C-27), 24.22 (C-22), 24.61 (C-6), 24.67 (C-26), 25.57 (C-29), 27.34 (C-2), 27.49 (C-23), 28.37 (C-21), 32.02 (C-12), 32.19 (C-19), 33.67 (C-28), 35.31 (C-10), 35.37 (C-20), 36.98 (C-1), 37.42 (C-15), 38.08 (C-13 or C-17), 38.09 (C-13 or C-17), 38.99 (C-4), 42.11 (C-14), 48.14 (C-9), 50.65 (C-5), 55.11 (C-18), 77.03 (C-16), 79.04 (C-3), 116.80 (C-7), 144.36 (C-8).

Bauerenol (2) Amorphous powder. $[\alpha]_D^{23}$ -21.7° ($c = 0.1$, CHCl_3); [lit.,⁴] $[\alpha]_D$ -25° (CHCl_3). IR $_{\text{max}}$ (CHCl_3) cm^{-1} : 3610, 3472. HR-MS m/z : 426.3852 (M^+ , Calcd for $\text{C}_{30}\text{H}_{50}\text{O}$; 426.3861). EI-MS m/z : 426 (M^+), 411 ($\text{M}^+ - \text{CH}_3$), 408 ($\text{M}^+ - 2\text{H}_2\text{O}$), 393 ($\text{M}^+ - \text{CH}_3 - \text{H}_2\text{O}$), 273, 259, 247, 229, 205. $^1\text{H-NMR}$ (CDCl_3 , 270MHz) δ : 0.75 (3H, s, H_3 -25), 0.86 (3H, s, H_3 -24), 0.90 (3H, d, $J=5.9\text{Hz}$, H_3 -30), 0.94 (3H, s, H_3 -27), 0.97 (3H, s, H_3 -23), 1.00 (3H, s, H_3 -26), 1.04 (3H, s, H_3 -28), 1.05 (3H, d, $J=5.9\text{Hz}$, H_3 -29), 3.24 (1H, m, H-3), 5.45 (1H, br s, H-7); Acetate: $^1\text{H-NMR}$ (CDCl_3 , 270MHz) δ : 0.77 (3H, s, H_3 -25), 0.85 (3H, s, H_3 -24), 0.91 (3H, d, $J=5.8\text{Hz}$, H_3 -30), 0.93 (3H, s, H_3 -27), 0.95 (3H, s, H_3 -23), 1.00 (3H, s, H_3 -26), 1.04 (3H, s, H_3 -28), 1.05 (3H, d, $J=5.9\text{Hz}$, H_3 -29), 2.05 (s, OAc), 4.52 (1H, dd, $J=11.1$, 4.8Hz, H-3), 5.41 (1H, br d, $J=4.0\text{Hz}$, H-7).

Isobauerenol (3) Amorphous powder. $[\alpha]_D^{22}$ +31.9° ($c = 0.2$, CHCl_3); [lit.,⁵] $[\alpha]_D$ +37.2° (CHCl_3). IR $_{\text{max}}$ (CHCl_3) cm^{-1} : 3609, 3474. HR-MS m/z : 426.3877 (M^+ , Calcd for $\text{C}_{30}\text{H}_{50}\text{O}$; 426.3861). EI-MS

m/z : 426 (M^+), 411 ($\text{M}^+ - \text{CH}_3$), 408 ($\text{M}^+ - 2\text{H}_2\text{O}$), 393 ($\text{M}^+ - \text{CH}_3 - \text{H}_2\text{O}$), 273, 259, 247, 229, 205. $^1\text{H-NMR}$ (CDCl_3 , 270MHz) δ : 0.80 (3H, s, H_3 -24), 0.88 (3H, s, H_3 -27), 0.90 (3H, d, $J=5.9\text{Hz}$, H_3 -30), 0.95 (3H, s, H_3 -25), 0.99 (3H, d, $J=6.3\text{Hz}$, H_3 -29), 1.00 (6H, s, H_3 -23, H_3 -26), 1.05 (3H, s, H_3 -28), 3.23 (1H, dd, $J=10.7$, 4.8Hz, H-3); Acetate: $^1\text{H-NMR}$ (CDCl_3 , 270MHz) δ : 0.84 (3H, s, H_3 -24), 0.87 (3H, s, H_3 -27), 0.88 (3H, s, H_3 -23), 0.90 (3H, d, $J=5.9\text{Hz}$, H_3 -30), 0.98 (3H, s, H_3 -25), 0.99 (3H, d, $J=6.9\text{Hz}$, H_3 -29), 1.00 (3H, s, H_3 -26), 1.05 (3H, s, H_3 -28), 2.05 (s, OAc), 4.50 (1H, dd, $J=11.4$, 4.8Hz, H-3).

REFERENCES

- 1) W. Shi, G. Han, *J. Chin. Pharm. Sci.*, **5**, 63 - 67 (1996).
- 2) N. Suzuki, M. Kikuchi, *Yakugaku Zasshi*, **112**, 571 - 576 (1992).
- 3) M. Kikuchi, N. Suzuki, *Chem. Pharm. Bull.*, **40**, 2753 - 2755 (1992).
- 4) P. Sengupta, H. N. Khastgir, *Tetrahedron*, **19**, 123 - 132 (1963).
- 5) S. K. Talapatra, S. Sengupta, B. Talapatra, *Tetrahedron Lett.*, 5963 - 5968 (1968).
- 6) K. Shiojima, Y. Arai, K. Masuda, Y. Takase, T. Ageta, H. Ageta, *Chem. Pharm. Bull.*, **40**, 1683 - 1690 (1992).