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BIOCHEMICAL STUDIES ON ETHYL METHANE SULFONATE-INDUCED CHLORENA MUTANT OF *TRITICUM DICOCCUM* VAR. KHAPLI

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SWAMINATHAN et al. (1) reported the occurrence of EMS (ethyl methane sulfonate)-induced chlorophyll deficient mutant of *Triticum dicoccum* var. Khapli. In view of the reports in literature (2-4) attributing the changes in morphological and physiological characteristics to the lack of one of the essential elements in the mutant, and describing the reversal, in some cases, caused by various external treatments, a number of experiments were performed in our laboratory, using the chlorena mutants. These are: (A) analysis of various elements in the leaves, viz. calcium, potassium, magnesium, phosphorus and iron, (B) foliar sprays of urea, copper, zinc, manganese, iron, boron and molybdenum, (C) placing the seedlings under long and short day conditions, and (D) placing the seedlings at different temperatures.

It was observed that there was no difference in concentrations of the elements analysed, except for phosphorus. In 30 day-old seedlings, its value is about 0.3% (on dry weight basis) while in normal plants values of 0.18% were observed. External treatments described above did not result in any recovery of the chlorophyll content of the leaves and the plants remained yellow in appearance.

In this communication, results of experiments on determinations of chlorophyll, carotenoids and total nitrogen in the leaves and various fractions of phosphorus in the chloroplasts of mutant and normal plants are reported.

Normal plants and M_2 generation of chlorena mutant (5) were raised in the field during 1965-66 and leaf samples of each were collected at 30 and 75 day stage. 30 Grams of leaves were cut into small pieces and homogenized in a Waring blendor with 100 ml portions of 0.3 M sodium chloride solution containing 0.02 M Tris buffer, pH 8.3 and 0.01 M sodium ascorbate. The homogenate was passed through muslin cloth and centrifuged at $40 \times g$ to remove the cell wall material. The superenatant was centrifuged at $1500 \times g$

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for 10 min. The residue was washed twice with the sodium chloride solution and centrifuged (6).

Various fractions of phosphorus in the chloroplast pellet were obtained by the method of SMILLIE and KROTKOV (7): Methanol-P fraction-extracted three times with 15 ml each of cold methanol containing 0.05 M formic acid. The fine suspension in each case is allowed to stand for a few minutes before centrfugation. Acid-P fraction—The residue thus obtained was extracted with 10 ml of 5 per cent trichloroacetic acid at 4°. Centifuged. Lipid-P fraction-The residue, after the isolation of the above fraction, was extracted with 15 ml lots of (i) 95 per cent ethanol (ii) ethanol: ether (1:1) and (iii) ether. For each extraction, the solvent was kept boiling for 20 sec. Nucleic acid-P fraction-The insoluble residue, after the above extraction, was dried, powdered and then extracted with 5 per cent perchloric acid at 90° for 15 min. centrifuged.

Phosphorus was estimated by following the method of KING (8). Extraction and estimation of chlorophylls and carotenoids were made according to the technique adopted by FRANK and KENNY (9). Total nitrogen was determined by KJELDAHL's method (10).

The contents of chlorophyll and carotenoids in the mutant were lower than in the normal plants. In the mutant, the ratio of the two pigments

Age (days)	Char in a	Chlorophyll	Carotenoids	Chlorophyll/	
	Strains	OD/mg, Fr. Wt. 660 mµ	0D/mg, Fr. Wt. 440 mµ	Carotenoids OD/OD	
		(10-4)	(10-4)		
30	Normal	14.5	44.5	1:3.0	
	Mutant	3.2	17.8	1:5.6	
75	Normal	12.5	38.7	1:3.1	
	Mutant	5.0	35.3	1:7.1	

TABLE I

Chlorophyll and carotenoid contents in normal and chlorena mutant

TABLE II

Analysis of chloroplast fractions of normal and mutant plants (Triticum dieoccum var. Khapli)

Age (days)	Strains	Dry wt. (%)	Chloroplast material						
			Dry Total wt. N		Alcohol-P	Acid-P	Lipid-P	Nucleic acid-P	Total P
			$\mu { m g}$	$\mu { m g}$	$\mu { m g}$	$\mu { m g}$	$\mu \mathbf{g}$	$\mu { m g}$	μg
30	Normal	16.3	431	244.2	6.93	7.30	2.83	6.27	23.33
	Mutant	12.5	315	191.5	3.50	15.50	4.70	5.88	29.58
75	Normal	20.1	628	261.5	8.02	11.44	2.89	8.66	31.01
	Mutant	19.2	443	206.3	4.88	14.89	6.97	8.22	34.96

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varied depending on the age of the plant. As compared with the normal chlorophyll: carotenoids ratio (OD/OD) of 1:3, the mutant showed ratios of 1:5.6 and 1:7.1 in 30 and 75 day-old seedlings, respectively (Table I). The dry weight (%) of the leaves in the mutant was less than that in the normal plant. The difference, however, diminished with the growth of the plants (see Table II), reflecting the recovery in chlorophyll content and photosynthetic activity in the older seedlings (Table I).

The results of analysis of various fractions of chloroplast material are summarized in Table II. A marked difference was observed only in acid soluble P, the mutant chloroplast showing two times as high a content of this fraction of P as compared with that in normal ones. Also in this case, the difference diminished with age of the plants.

More detailed characterization of the non-colored material in the mutant and normal chloroplasts of *Triticum dicoccum* needs further investigation of the individual components of the plastids.

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REFERENCES

- (1) M. S. SWAMINATHAN, V. L. CHOPRA and S. BHASKARAN. 1962. Chromosome aberrations and the frequency and spectrum of mutations induced by ethyl methane sulfonate in barley and wheat. *Ind. Jour. Gen. & Plant Breeding*, 22, 192-207.
- (2) M. G. WEISS. 1943. Inheritance and physiology of efficiency in iron utilization in Soybean. *Genetics*, 28, 253-268.
- (3) B. KESSLER. 1958. Further evidence on the effect of bicarbonate ions upon nucleic acid metabolism and ion uptake in fruit trees susceptible and nonsusceptible to lime-induced chlorosis. KTAVIM, 8, 287-293.
- (4) W. D. BELL, L. BOGORAD and W. J. MCILRATH. 1963. Yellow-stripe phenotype in maize I. Effects of ys1 locus on uptake and utilization of iron. Botanical Gaz., 124, 1-5.
- (5) V. L. CHOPRA and M. S. SWAMINATHAN. 1966. Mutagenic efficiency of individual and combined treatments of ethyl methane sulfonate and hydroxyl amine in emmer wheat. *Ind. Jour. Gen. & Plant Breeding*, 29, 59-62.
- (6) S. HAQ and W. Z. HASSID. 1965. Biosynthesis of sucrose phosphates in sugarcane leaf chloroplast. *Plant Physiol.*, 4, 591-599.
- (7) R. M. SMILLIE and G. KROTKOV. 1960. Estimation of nucleic acids in some algae and higher plants. *Can. Jour. Bot.*, 38, 31-49.
- (8) E. J. KING. 1932. The colorimetric determination of phosphorus. Biochem. Jour., 26, 292-297.
- (9) S. FRANK and A. L. KENNY, 1955. Chlorophyll and carotenoids destruction in the absence of light in seedlings of Zea mays. Plant Physiol., 30, 413-418.
- (10) W. HERWITZ (ED.). 1960. Methods of Analysis. A. O. A. C., 9th edition, p 643.