

Spatio-Temporal Pattern of DNA Synthesis Detected by Bromodeoxyuridine Labeling in the Mouse Endometrial Stroma during Decidualization

NAOSHI OHTA^{1,2}, TAKAO MORI¹, SEIICHIRO KAWASHIMA¹,
SHINOBU SAKAMOTO³, HIDESHI KOBAYASHI²

¹*Zoological Institute, School of Science, University of Tokyo, Bunkyo-ku, Tokyo 113,*

²*Research Laboratory, Zenyaku Kogyo Co., Ltd. Nerima-ku, Tokyo 178,*

³*Department of Endocrinology, Medical Research Institute,
Tokyo Medical and Dental University, Bunkyo-ku,
Tokyo 113, Japan*

ABSTRACT—In order to examine the patterns of proliferation and differentiation of endometrial stromal cells before and during decidualization in pseudopregnant mice, the rate of DNA synthesis was immunocytochemically determined by means of bromodeoxyuridine (BrdU) labeling. On day 4 of pseudopregnancy induced by mating with vasectomized male, both uterine horns were traumatized to induce deciduoma. On day 5 of pseudopregnancy (one day after traumatization), BrdU-labeling index was markedly increased, and the labeled cells were found in almost all parts of endometrial stroma. From day 6 to day 8 of pseudopregnancy (2–4 days after traumatization), the labeling index remained high in the stromal cells of all parts except for the periluminal region. In the endometrial stromal cells in the peripheral region of myometrium, however, the labeling index was maximum on day 8 and decreased remarkably on day 9. In the stromal cells in the periluminal region where decidual cells developed, the labeling index was high on day 5 and low on day 6, no labeled cells being found on day 8. These results clearly show that each region of uterine endometrial stroma has a different responsiveness to traumatization, and each region plays a different role in the formation of deciduoma.

INTRODUCTION

Immunohistochemical detection of bromodeoxyuridine (BrdU), which is a uridine analogue and incorporated selectively into the cellular DNA at S-phase of the cell cycle, has been proven useful for the analysis of cell proliferation in place of ³H-thymidine incorporation into replicating cells [5].

Differentiation of the endometrial stromal cells into decidual cells occurs soon after the implantation of blastocysts. In mice and rats, however, decidual reaction can be induced artificially in the uteri without blastocysts by mechanically scratching the luminal surface [17]. Changes in the structure and function of uterine tissue during decidualization are mainly controlled by the ovarian estrogen and progesterone [2, 11, 13, 14]. Decidualization is a highly regulated process characterized by a variety of events including increase in DNA synthesis [1, 8, 15], changes in vascular permeability [7], and polyploidization and hypertrophy of stromal cells [12, 16]. Therefore, formation of deciduoma has been widely applied as a useful experimental model for the study not only of implantation but also of the mechanisms of cell proliferation and differentiation. BrdU labeling patterns of the cells in uterine tissue were reported in normal cycling and prepubertal mice [6].

Because the changes of cell proliferation during decidualization as a function of time have not been reported, the present study was designed to examine the spatial and

temporal patterns of DNA synthesis in the mouse uterine stromal cells during decidualization after traumatization.

MATERIALS AND METHODS

Animals

Female mice of the ICR strain purchased from Japan CLEA Inc. (Tokyo, Japan) were used in the present study. They were housed in plastic cages (3–7 mice per cage) under controlled lighting (12-hr light and 12-hr darkness; lights on at 06:00) and temperature (25 ± 0.5°C), and were provided with a commercial diet (CE-7: Japan CLEA) and tap water *ad libitum*.

Induction of deciduoma

Virgin female mice at 50–60 days of age were mated with vasectomized males to induce pseudopregnancy. The day when a vaginal plug was found was designated as day 1 of pseudopregnancy. On day 4 of pseudopregnancy, the anti-mesometrial luminal surface in both uterine horns was traumatized by a single scratch with a bent needle. The needle was inserted into the uterine lumen from a small incision made with a small scissors at the posterior end of uterine horn, adjacent to the uterine cervix, under light nembutal anesthesia [17]. The pseudopregnant mice were killed by cervical dislocation on various days after traumatization (Fig. 1). In order to check the effect of trauma, some pseudopregnant mice without traumatization were killed as controls between 2 and 10 days after mating with vasectomized males. Immediately after autopsy, the uterine horns were removed and weighed. The uterine weight was used as a parameter of decidual reaction.

In addition, virgin cycling mice at 50–60 days of age were also killed at various phases of the estrous cycle and the uterine weights were recorded.

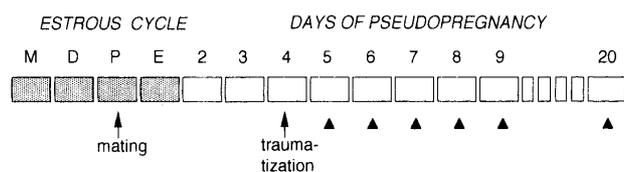


FIG. 1. Experimental schedule. Pseudopregnant mice received a decidual stimulus (traumatization) on day 4 of pseudopregnancy and were given a single injection of BrdU on various days (\blacktriangle) after traumatization. M: metestrus, D: diestrus, P: proestrus, E: estrus.

BrdU labeling and immunocytochemistry

The pseudopregnant mice received a single intravenous injection of bromodeoxyuridine (BrdU, 30 mg/kg body weight: Amersham, UK) at 24-hr intervals after traumatization. Four hr after BrdU injection, the uteri were fixed in ice cold 10% phosphate-buffered neutral formalin for 5 hr at room temperature. The uteri were dehydrated, embedded in paraffin, and the sections were cut at $4\mu\text{m}$ thickness.

After deparaffinization, the sections were washed in 0.01M phosphate-buffered saline (PBS, pH 7.4) three times and digested with 0.1% trypsin (Sigma) in 0.1% CaCl_2 (pH 7.8) for 20 min at 37°C . After washing in PBS (15 min, 3 times), endogenous peroxidase activity was blocked by immersing the sections in 0.3% H_2O_2 in methanol for 20 min, followed by washing in PBS (15 min, 3 times). Thereafter, the sections were incubated with monoclonal anti-BrdU antibody containing 10 units/ml nuclease (Amersham) for 1 hr at room temperature and then rinsed in PBS (15 min, 3 times). Finally, the sections were incubated with peroxidase-conjugated rabbit anti-mouse IgG for 30 min at room temperature. After washing in PBS (15 min, 3 times), the antibody binding sites were visualized by 0.05% 3, 3'-diaminobenzidine tetrahydrochloride solution. Each incubation was conducted in a moist chamber at room temperature. After immunostaining, the sections were counterstained with 0.1% Kernechtrot in 5% $\text{Al}_2(\text{SO}_4)_3$, dehydrated through an ethanol series, cleared in xylene, and mounted. The immunocytochemistry was controlled by sections overlaid with PBS instead of anti-BrdU antibody, which showed no immunoreactivity.

Measurement of labeling index

In order to examine the BrdU labeling index, two sections which were separated by at least $40\mu\text{m}$ apart were randomly chosen from the middle part of the uterine horn or the middle part of deciduoma in each mouse. Cell counting was carried out in the four regions; anti-mesometrial side of the endometrial stroma (AME), periluminal endometrial stroma (PLE), peripheral endometrium adjacent to the myometrium (PPE), and mesometrial side of the endometrial stroma (MME) (Fig. 2). Total number of BrdU labeled cells was counted out of 1,000 cells each in two sections from the four regions by using

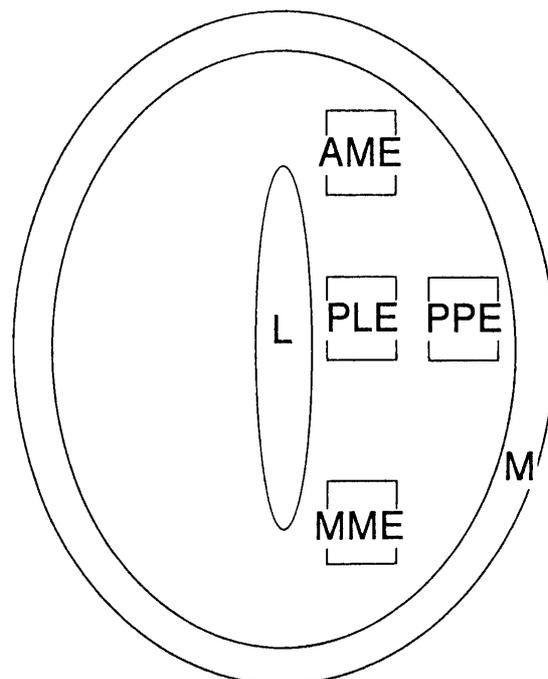


FIG. 2. Four regions of uterus for examining BrdU labeling index. L: lumen, M: myometrium, AME: antimesometrial side of endometrial stroma, PLE: periluminal endometrial stroma, PPE: peripheral endometrial stroma adjacent to myometrium, MME: mesometrial side of endometrial stroma.

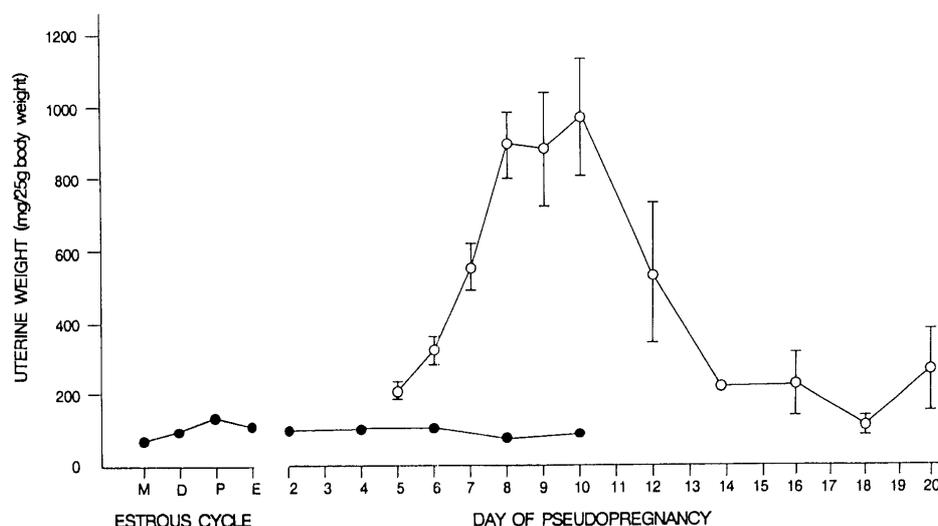


FIG. 3. Changes in uterine weights in mice during estrous cycle and pseudopregnancy with (\circ -) or without (\bullet -) traumatization. Deciduoma were induced on the next day of traumatization given on day 4 of pseudopregnancy. Each point depicts the mean and SEM of 3-5 mice. M: metestrus, D: diestrus, P: proestrus, E: estrus.

an image processor-analyzer (LUZEX; NIRECO Co. Ltd, Tokyo). The labeled indices were expressed as percentages of labeled cells per 1,000 cells.

Statistical analysis

The statistical significance of the difference between groups were evaluated by one-way analysis of variance (ANOVA), followed by Duncan's multiple range test for the uterine weights and labeled indices.

RESULTS

Changes in uterine weight during decidualization

Changes in uterine weights during the estrous cycle and pseudopregnancy before and after traumatization are shown in Figure 3. In normal cycling mice, the uterine weight was lowest at metestrus, followed by an increase during diestrus ($P < 0.01$). The weight reached maximum at proestrus, the value being significantly higher than those in the other phases of estrous cycle ($P < 0.01$).

The uterine weight of pseudopregnant mice without traumatization between 2 and 10 days after the mating was almost the same as that of diestrous mice. Traumatization of the uteri on day 4 of pseudopregnancy resulted in the development of deciduoma in the next day. The weight increased rapidly after traumatization, reaching maximum on

day 8 of pseudopregnancy, and decreased on day 12 and onward. The weights were significantly higher between days 6 and 12 than the other stages of pseudopregnancy with traumatization ($P < 0.01$, in all comparisons).

BrdU labeling index

On day 2 of pseudopregnancy before traumatization, BrdU labeled cells were observed in the luminal and glandular epithelia but not in the endometrial stroma. On days 3 and 5 of pseudopregnancy before and one day after traumatization, a large number of labeled cells were observed in the endometrial stroma but very rarely in the luminal epithelium (Fig. 4a). On days 7 and 8 of pseudopregnancy given no traumatization, labeled cells were not found in the stroma and appeared again in the luminal epithelium.

The BrdU-positive cells on day 5 of pseudopregnancy in mice given traumatization (Fig. 4b) were more numerous compared to those on day 3 of pseudopregnancy (Fig. 4a) and on day 5 of pseudopregnancy without traumatization (data not shown, but regardless of the regions, the indices were less than 1% in mice given no traumatization).

Detailed spatio-temporal patterns of BrdU labeling index during decidualization are shown in Figure 5. On day 5 through 8 of pseudopregnancy with traumatization, the indices were significantly higher in the endometrial stroma than

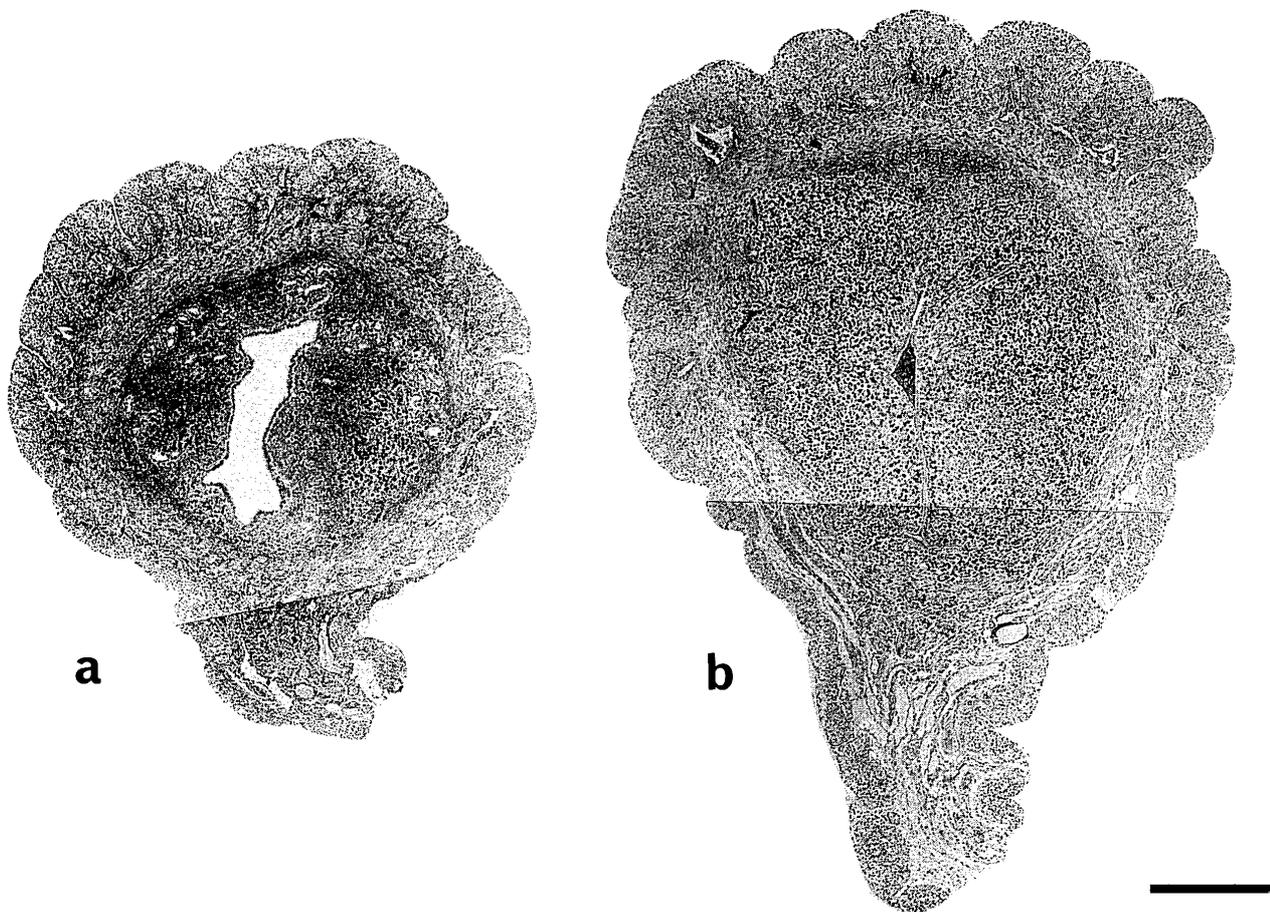


FIG. 4. Uteri of mice on day 3 (a) and day 5 (b) of pseudopregnancy, one day before and after traumatization, respectively. BrdU-labeled cells (black dots) were visible in the endometrial stroma. Bar: 500 μ m

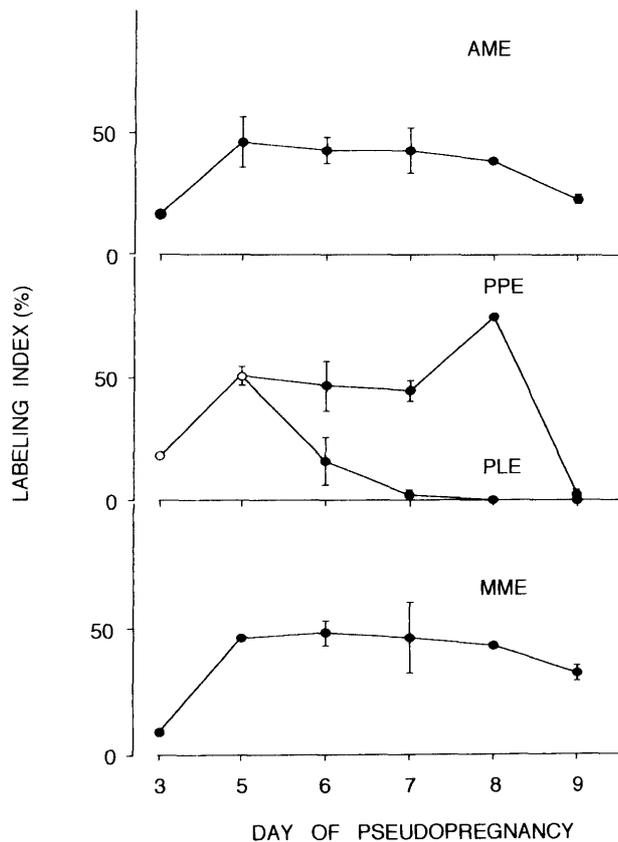


FIG. 5. Changes of BrdU labeling index in the endometrial stromal cells during decidualization. Each point depicts the mean and SEM of 3–5 mice. AME: antimesometrial side of endometrial stroma, PPE: peripheral endometrial stroma adjacent to myometrium, PLE: periluminal endometrial stroma, MME: mesometrial side of endometrial stroma. On days 3 and 5, as PPE and PLE could not be counted separately, pooled data are shown.

those on day 3 at 0.01 level, except for PLE (Figs. 5–7). On day 6 of pseudopregnancy with traumatization, the plump cells with large nuclei over $25\mu\text{m}$ in diameter appeared in the PLE and AME (Fig. 8). In PLE, the percentage of labeled stromal cells on day 5 was significantly higher than that in the other stages, respectively ($P < 0.01$). The percentage reduced rapidly on day 6 and BrdU labeling was no more detected on day 8 (Fig. 5). On day 8, labeling indices tended to be lower in the regions of the endometrium except for PPE. On day 9, all labeling indices were almost the same as those on day 3 except for MME (Figs. 5 and 9). Many degenerating cells with pyknotic nuclei appeared on days 8 and 9 of pseudopregnancy with traumatization.

DISCUSSION

Cell proliferation and differentiation during decidualization in pregnant [3] or pseudopregnant rodents [4, 9, 17] have extensively been studied. It is well known that the differentiation of endometrial stromal cells to decidual cells occurs in response to the implantation of blastocysts or traumatization of artificial stimuli. In mice, the sensitivity of the uterus to a decidualogenic stimulus is known to be the

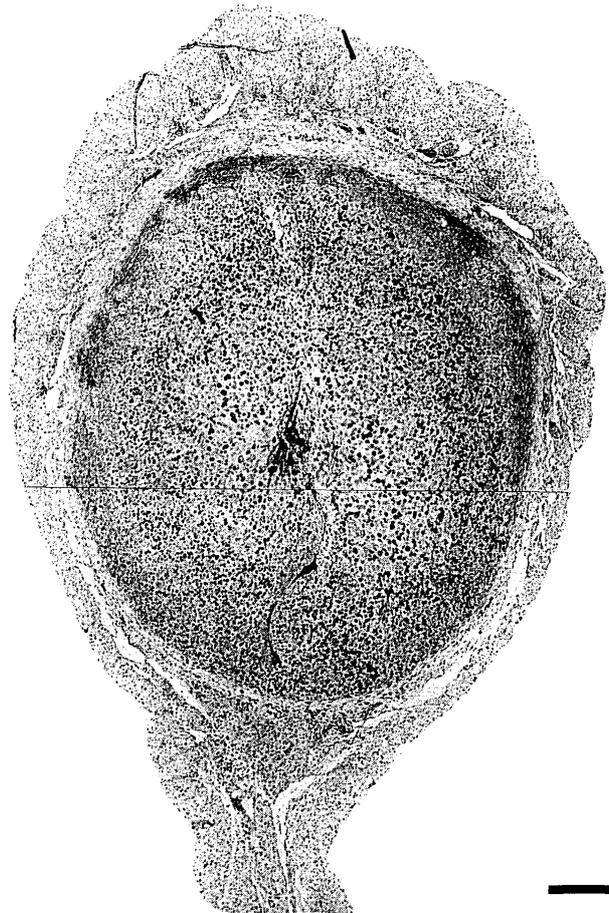


FIG. 6. Uterus of a mouse on day 6 of pseudopregnancy 2 days after traumatization. BrdU-labeled cells were present in almost all regions of the endometrial stroma except for PLE. Bar: $500\mu\text{m}$

highest on day 4 of pseudopregnancy [2, 10]. In the present study, the uterine weight increased immediately after traumatization on day 4 and reached maximum on day 10 of pseudopregnancy. The weight markedly decreased on day 12 and returned to the normal diestrous level on day 18. These findings accord well with the previous results in rats [17].

In the present study, DNA synthesis was detectable by the presence of BrdU-labeled cells in the luminal epithelial cells on day 2 of pseudopregnancy. On day 3 of pseudopregnancy, some stromal cells in the endometrium began to show DNA synthesis. If decidualogenic stimuli were not given to the uterus, the activity of DNA synthesis in the stromal cells decreased within a few days. These findings may reflect that the stromal cells are ready to respond to decidualogenic stimuli on day 3 of pseudopregnancy. On day 5 of pseudopregnancy with traumatization, the labeled cells were present extensively and evenly in all the four regions of endometrial stroma. Thereafter, BrdU-labeled cells greatly decreased in the PLE on days 6 and 7 of pseudopregnancy. Ledford *et al.* [8] have stated that in mice rapid cell proliferation begins approximately 30 hr after decidualogenic stimulation and continued for 72 hr in the endometrial stroma. After the initiation of decidualization, however, a population of stromal cells

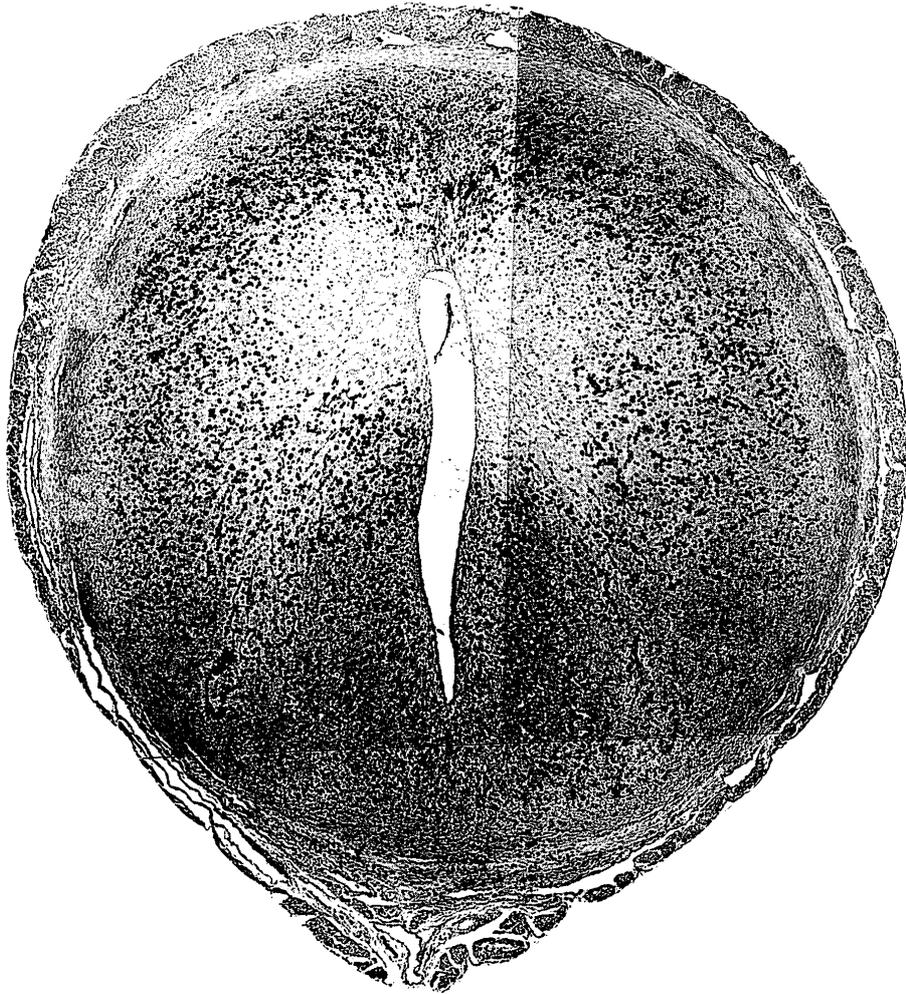


FIG. 7. Uterus of a mouse on day 7 of pseudopregnancy 3 days after traumatization. Only a few BrdU-labeled cells were visible in PLE. Bar: 500 μ m

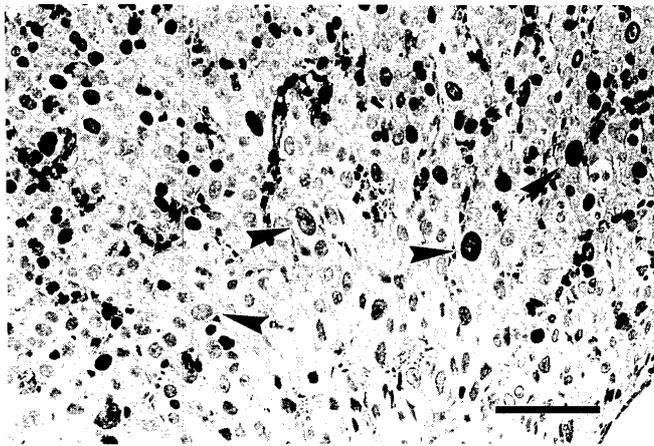


FIG. 8. Antimesometrial side of endometrial stroma in a mouse on day 6 of pseudopregnancy 2 days after traumatization. Plump cells with large nuclei (arrowheads) appeared. Bar: 100 μ m

is known to synthesize DNA and differentiate into polyploid decidual cells without cell division [1]. Deciduomal cells called plump cells in the present study are distributed exclusively in the periluminal part of endometrial stroma where the implantation normally occurs. Thus, it seems likely that the proliferation of decidual cells ceases and differentiation begins on day 6 or 7 of pseudopregnancy (2–3 days after traumatization/implantation).

During decidualization, a remarkable rise of DNA synthesis in peripheral endometrial stroma adjacent to the myometrium (PPE) occurred between day 6 and day 8 of pseudopregnancy. Proliferated stromal cells in this region may contribute to the reconstruction of endometrial tissue after the regression of preformed decidual tissue. It is known that the life span of the rat decidualoma is limited and frequent cell death occurs on day 9 of pseudopregnancy [14]. The present findings clearly show that regression of decidualoma begins in most parts of the endometrium from day 8 of pseudopregnancy, because many degenerated cells were encountered on days 8 and 9 of pseudopregnancy.



FIG. 9. Uterus of a mouse on day 9 of pseudopregnancy 5 days after traumatization. The number of BrdU-labeled cells became decreased. Bar: 500 μ m

The present findings demonstrated that activity of DNA synthesis in the endometrium varies during decidualization in several regions of the endometrial stroma. This implies that each region has a different responsiveness to traumatization and that it plays a different role during the development of deciduoma.

ACKNOWLEDGMENTS

The authors would like to express our cordial thanks to Dr. M. K. Park for his advice and encouragement during this study. This study was supported by Grants-in-Aid for Scientific Research from the Ministry of Education, Science and Culture, Japan to T. M., S. K. and S. S., respectively and a Research Grant from Zenyaku Kogyo,

Ltd. to S. K.

REFERENCES

- 1 Das RM, Martin L (1978) Uterine DNA synthesis and cell proliferation during early decidualization induced by oil in mice. *J Reprod Fert* 53: 125-128
- 2 Finn CA (1966) Endocrine control of endometrial sensitivity during the induction of the decidual cell reaction in the mouse. *J Endocr* 36: 239-248
- 3 Finn CA, Martin L (1967) Patterns of cell division in the mouse uterus during early pregnancy. *J Endocr* 39: 593-597
- 4 Galassi L (1968) Autoradiographic study of the decidual cell reaction in the rat. *Dev Biol* 17: 75-84
- 5 Gratzner HG (1982) Monoclonal antibody to 5-bromo- and

- 5-iododeoxyuridine: a new reagent for detection of DNA replication. *Science* 218: 474-475
- 6 Hanazono M, Yoshiki A, Ota K, Kitoh J, Kusakabe M (1990) DNA replication in uterine cells of adult and prepubertal mice under normal and hormonally stimulated conditions detected by bromodeoxyuridine labeling method. *Endocrinol Japon* 37: 183-191
 - 7 Kennedy TG (1979) Prostaglandins and increased endometrial vascular permeability resulting from the application of an artificial stimulus to the uterus of the rat sensitized for the decidual cell reaction. *Biol Reprod* 20: 560-566
 - 8 Ledford BE, Rankin JC, Froble VL, Serra MJ, Markwald RR, Baggett B (1978) The decidual cell reaction in the mouse uterus: DNA synthesis and autoradiographic analysis of responsive cells. *Biol Reprod* 18: 506-509
 - 9 Marcus GJ (1974) Mitosis in the rat uterus during the estrous cycle, early pregnancy, and early pseudopregnancy. *Biol Reprod* 10: 447-452
 - 10 Martin L, Finn CA (1968) Hormonal regulation of cell division in epithelial and connective tissues of the mouse uterus. *J Endocr* 41: 363-371
 - 11 Moulton BC, Blaha GC (1978) Separation of decidual cells by velocity sedimentation at unit gravity. *Biol Reprod* 18: 141-147
 - 12 Moulton BC, Koenig BB (1984) Uterine deoxyribonucleic acid synthesis during preimplantation in precursors of stromal cell differentiation during decidualization. *Endocrinology* 115: 1302-1307
 - 13 Ohta Y (1991) Deciduoma formation in pseudopregnant rats bearing pituitary grafts. *Zool Sci* 8: 75-80
 - 14 O'shea JD, Kleinfeld RG, Morrow HA (1983) Ultrastructure of decidualization in the pseudopregnant rat. *Am J Anat* 166: 271-298
 - 15 Tachi C, Tachi S, Lindner HR (1972) Modification by progesterone of oestradiol-induced cell proliferation, RNA synthesis and oestradiol distribution in the rat uterus. *J Reprod Fertil* 31: 59-76
 - 16 Takewaki K (1969) Formation of decidualomata in immature rats with luteinized ovaries. *Annot Zool Japon* 42: 126-132
 - 17 Velardo JT, Dawson AB, Olsen AG, Hisaw FL (1953) Sequence of histological changes in the uterus and vagina of the rat during prolongation of pseudopregnancy associated with the presence of decidualomata. *Am J Anat* 93: 273-305